ABSTRACT
Background Spread of SARS-CoV-2 by aerosol is considered an important mode of transmission over distances >2 m, particularly indoors.
Objectives We determined whether SARS-CoV-2 could be detected in the air of enclosed/semi-enclosed public spaces.
Methods and analysis Between March 2021 and December 2021 during the easing of COVID-19 pandemic restrictions after a period of lockdown, we used total suspended and size-segregated particulate matter (PM) samplers for the detection of SARS-CoV-2 in hospitals wards and waiting areas, on public transport, in a university campus and in a primary school in West London.
Results We collected 207 samples, of which 20 (9.7%) were positive for SARS-CoV-2 using quantitative PCR. Positive samples were collected from hospital patient waiting areas, from hospital wards treating patients with COVID-19 using stationary samplers and from train carriages in London underground using personal samplers. Mean virus concentrations varied between 429 500 copies/m³ in the hospital emergency waiting area and the more frequent 164 000 copies/m³ found in other areas. There were more frequent positive samples from PM samplers in the PM2.5 fractions compared with PM10 and PM1. Culture on vero cells of all collected samples gave negative results.
Conclusion During a period of partial opening during the COVID-19 pandemic in London, we detected SARS-CoV-2 RNA in the air of hospital waiting areas and wards and of London underground train carriage. More research is needed to determine the transmission potential of SARS-CoV-2 detected in the air.

INTRODUCTION
COVID-19 is an acute respiratory disease caused by the novel SARS-CoV-2. Since its identification in Wuhan, China, in 2019, SARS-CoV-2 has infected 434 million people and caused 5.9 million deaths worldwide, as of February 2022 (WHO, 2022). Improving our understanding of the characteristics and behaviour of this coronavirus which make it highly transmissible is key to developing future mitigation measures to limit its transmission.
aerosols generated by an infected singer during a choir practice transmitted the infection to 32 others in the vicinity.

Under controlled temperature and humidity conditions, aerosolised SARS-CoV-2 is viable and retains infectivity for 3–16 hours. Clusters of SARS-CoV-2 RNA have been reported in aerosols collected from air samples in multiple indoor and outdoor settings, and studies by Lednicky et al captured viable SARS-CoV2 virus in air samples by culture on Vero E6 cells. Therefore, we (i) characterised the distribution of SARS-CoV-2 on airborne particles in hospital and other public environments, (ii) assessed the infectivity of SARS-CoV-2 collected from air samples and (iii) compared the performance of a range of air samplers that collect total or size-fractionated particulates.

**METHODS**

**Sampling sites**

Air samples were collected between 10 March 2021 and 14 December 2021 starting from four major hospitals (Charing Cross Hospital, Chelsea and Westminster Hospital, Royal Brompton Hospital and the Royal Marsden Hospital) in intensive care units (ICUs) with adults suffering from COVID-19, respiratory wards and public waiting areas. Collection in ICUs was to test the likelihood of collecting the virus from the air particularly when there was patients with COVID-19 being treated in these rooms. As lockdown measures were gradually lifted after the third national lockdown in England which commenced on 6 January 2021, we sampled in public or semi-public settings from Paddington Rail station, Paddington Underground station (Bakerloo line), inside London Underground carriages, Imperial College University campus and Hampstead Garden Suburb Primary School. Sampling frequency was based on obtaining permissions and being granted access to the sampling locations while sampling frequency was dependent on sampling device used and the capacity to supervise sampling equipment where necessary.

**Air sampling instruments and postsampling processing**

A range of sampling instruments was used to collect air particles and liquid bioaerosols (table 1). These included liquid-based total suspended particulate (TSP) samplers, size-segregated particulate matter (PM) samplers and also portable samplers. All samples were stored in Petri dishes and transported on ice to a containment level 2 (CL2) laboratory for processing. The sampling instruments and the processing of the polyurethane foam, Teflon and gelatin filters are described in the online supplemental file 1.

**RNA extraction and RT-qPCR for SARS-CoV2**

Viral RNA was extracted from 140 µL of sample solution using the QIAamp Viral RNA mini-Kit (Qiagen, Hilden, Germany), following manufacturer’s instructions, and eluted in 35 µL. Extracted viral RNA from each sample underwent RT-qPCR targeting the n-gene (table 2).

To calculate the viral copy number, we simultaneously ran an eight-fold serial dilution of RNA extracted from the research reagent for SARS-CoV-2 RNA (National Institute for Biological Standards and Controls (NIBSC) 19/304) alongside RNA extracted from environmental air samples in a qPCR reaction targeting the N-gene of SARS-CoV-2, with RNase P as an internal control. All samples were run in duplicate. We then calculated the viral copy number per qPCR reaction and subsequently copy number per nanogram of RNA in environmental air samples. Viral copy number (log10) was estimated from Ct values of environmental air samples using the standard curve generated from the qPCR, the research reagent for SARS-CoV-2 RNA (NIBSC 19/304). This value was antilogged and the average divided by the total RNA per qPCR reaction to give the SARS-CoV-2 copy number per nanogram of RNA. The following equation was used to calculate the total virus copy number: virus copy number per ng RNA x total RNA per sample.

The concentration of virus in the air was calculated by dividing the estimated total virus copy number by the total volume of air sampled per cubic metre and expressed as virus copy number per m³. The volume of air sampled was calculated by multiplying the sampling rate with the sampling time.

All samples were run in duplicate alongside nuclease-free water as a non-template control and an NIBSC standard 19/304 as a positive control.

**Culture on Vero E6 cells**

Vero E6 cells were cultured in a containment level 3 laboratory at 37°C/5% CO₂ in Dulbecco’s modified eagle medium (DMEM) with 10% fetal bovine serum in T75 flasks. Once at 70% confluency, cultured Vero E6 cells were added to a 24-well plate (20000 cells/well) and inoculated with extracted air samples in a limiting dilution, with the addition of DMEM supplemented with 8x penicillin, streptomycin and amphotericin B. Culture plates were incubated at 37°C/5% CO₂ for a maximum of 6 days and assessed every 24 hours for cytopathic effects using a microscope. If disruption to the cell monolayer was observed during the incubation period, the relevant well was scraped and 100 µL of cells and media placed in 300 µL DNA/RNA shield to inactivate the sample. The samples were then extracted using the Viral Maghead Kit protocol on the OPentrons robot and 1 µL of template tested for the N1 and E gene using qPCR (Roche Master Hydrolisis Probes kit and the Roche 480 Lightcycler machine) performed according to manufacturer’s instructions.
Patient and public involvement

There has been no patient or public involvement.

RESULTS

In total, 207 samples were collected across all sites over 8 months using eight sampling instruments (table 3). Of these, 20 (9.7%) samples tested positive for detectable SARS-CoV-2 RNA, as summarised in table 4. Most of the positive samples were collected in hospital environments (n=15), from ‘lower-risk’ publicly accessible communal waiting areas (n=10) and from ‘higher-risk’ hospital rooms with a confirmed SARS-CoV-2-positive in-patient on ICUs and respiratory wards (n=5). The remaining five positive samples were from a London Underground train carriage, collected using a portable sampler on two separate London Underground journeys. The remaining 187 samples were negative on PCR testing. Figure 1 summarises the total number of collected samples at each site,
using each instrument, together with the SARS-CoV-2-positive samples. The only situation where SARS-CoV2 infection was confirmed was in the patients treated in ICUs. We did not check for the presence of SARS-CoV2 infection in any of the other sites where we sampled the air.

**Hospital wards**

One hundred and thirteen samples were collected across four hospitals in ICUs, respiratory wards and communal waiting areas. Forty samples were collected in negative pressure rooms in the ICU in two hospitals, 8 using PM samplers (mini volume sampler (MVS) and PDR10) and 32 using TSP collectors (Sartorius MD8, gelatin-loaded button sampler and Biospot VIVAS). All three positive samples were collected in the same sampling session using the MVS, PDR10 and Biospot VIVAS, in the room of a conscious patient breathing high-flow oxygen.

A total of 11 samples were obtained from respiratory wards using TSP collectors (MD8, Biospot VIVAS and gelatin-loaded SKC Button sampler), including one negative pressure room and two ambient pressure rooms. Two positive samples were obtained from two different locations, one in an ambient pressure room using the gelatin-loaded SKC Button sampler, with the other obtained in a negative pressure room using the Biospot VIVAS. Although both rooms were from different hospital sites, they both housed self-ventilating patients wearing a mask for supplementary oxygen.

**Communal hospital areas**

Sixty-two samples were collected from four communal waiting areas across four hospitals. Most samples (60) were collected using size-fractionated PM samplers (Harvard impactors, MVS, PDR10) and 2 from TSP collectors. Twenty-three samples were collected in emergency department (ED) ED waiting rooms, 26 in an outpatient waiting area and 13 in a chemotherapy day unit (CDU). Ten samples were positive in total, eight from two separate sampling sessions in the CDU and two from a general hospital ED waiting area. All positive samples were collected using size-fractionated PM samplers.

**London Underground and railway station**

Sixty-nine samples were collected using the Harvard impactors, cascade impactor, MVS, PDR and VIVAS from high-footfall areas of public transport including 18 from a major railway station, 26 in the ticket hall, near escalators and on the platform of a London Underground station and 25 from inside a deep-line London Underground train carriage using a portable sampling pump attached to polytetrafluoroethylene (PTFE)-loaded filter cassettes. All samples collected from the railway and underground stations were negative on PCR. Of the 25 samples taken inside the train carriage, 5 were positive for SARS-CoV2 RNA. These were collected from two separate journeys with the personal sampler worn by the same person.

All 28 samples taken from a university campus, including 11 from a busy university bar and 7 from a university engineering workshop were negative. All six samples collected in a primary school were also negative.

**Airborne virus concentrations in different environments**

Mean virus concentrations across different locations were expressed as virus copies/m³ (figure 2). The amount of virus collected across all samplers ranged from 118 virus copies/m³ of air to 707 284 copies/m³, with the higher concentrations of virus surpassing what has been reported previously (table 5). In hospital public areas, these were highest in the ED waiting area (429 500 copies/m³) and were markedly higher than mean concentrations measured in the CDU (75 523 copies/m³). Moreover, mean virus concentrations were lower on the respiratory ward cubicles: 539 copies/m³ in a respiratory ward negative pressure cube and 78 850 copies/m³ in an ICU negative pressure cube. All measurements conducted in hospital cubicles involved a self-ventilating patient who was within 1 week of admission to hospital and had a positive SARS-CoV-2 PCR test within 24 hours of sampling. The second highest mean concentration of virus (119 560 copies/m³) was measured inside the London Underground train carriage.

Two samples, both collected from the CDU from two separate devices in the same sampling session, underwent successful genomic sequencing to determine the SARS-CoV2 variant (see table 4). Both were related to the

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**Table 2** SARS-CoV-2 primers used for detection of SARS-CoV-2 RNA by RT-qPCR

<table>
<thead>
<tr>
<th>Oligonucleotide</th>
<th>Sequence 5’&gt;3’ (position)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>N gene Taq1</td>
<td>TCTGTAAAGGCAACACCAAA (28992)</td>
<td>HSL</td>
</tr>
<tr>
<td>N gene Taq2</td>
<td>TGTATGCTTTAAGTCGAGTAC (29073)</td>
<td>HSL</td>
</tr>
<tr>
<td>N gene Probe</td>
<td>(6FAM)CTGTCAGAAGATGCTGAGGC (BHQ1) (29023)</td>
<td>HSL</td>
</tr>
<tr>
<td>RNaseP Taq1</td>
<td>AGATTGGAACGTGCGGCGG</td>
<td>Emery et al⁶⁶</td>
</tr>
<tr>
<td>RNaseP Taq2</td>
<td>GAGCGGCGCTGTCTCCACCGT</td>
<td>Emery et al⁶⁶</td>
</tr>
<tr>
<td>RNaseP Probe</td>
<td>(Cyanine5)TTCTGACCCTGAGGC (28992)</td>
<td>Emery et al⁶⁶</td>
</tr>
</tbody>
</table>

HSL, Health Services Laboratories; RT-qPCR, reverse transcription quantitative PCR.
B1.1.7 variant, with one sample demonstrating a spike-protein mutation associated with S-variant SARS-CoV2 lineage B1.1.7.

**Size distribution of particles with positive samples**

SARS-CoV-2 RNA was most frequently detected in PM samplers compared with liquid-based TSP samplers (PM: 12 samples (85%) vs liquid-based TSP: 8 samples (15%)) across all settings (figure 3A). Positive samples collected on PM filters were more frequent in the PM2.5 size fraction (n=10). Using the Harvard sampler that fractionated the particles, two positive samples were detected on PM10 fractions on two different occasions, with positive samples also simultaneously detected in the PM2.5 fraction on both occasions with the PM10 fraction containing a higher amount of virus compared with the PM2.5 fraction. The VIVAS collected the most liquid-based positive samples (n=2) with the only other positive sample coming from a gelatin-loaded button sampler.

### Table 3

<table>
<thead>
<tr>
<th>PM samplers</th>
<th>TSP</th>
<th>MD8</th>
<th>Sartorius</th>
<th>Biospot</th>
<th>VIVAS</th>
<th>SKC</th>
<th>Coriolis</th>
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<tbody>
<tr>
<td>Number of samples</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
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<td>2</td>
<td>3</td>
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<td>0</td>
<td>1</td>
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<td>0</td>
<td>0</td>
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<td>0</td>
<td>0</td>
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<td>2</td>
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<tr>
<td>Chelsea and Westminster Hospital emergency department waiting room</td>
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<td>8</td>
<td>0</td>
<td>2</td>
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<td>0</td>
<td>0</td>
<td>0</td>
<td>10</td>
<td>3</td>
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<td>Royal Marsden Oncology medical day unit</td>
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<td>8</td>
<td>0</td>
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<td>3</td>
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<td>0</td>
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<td>0</td>
<td>2</td>
<td>0</td>
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<td>Paddington Underground escalators</td>
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<td>0</td>
<td>2</td>
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<td>Paddington Underground platform (Bakerloo)</td>
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<td>London Underground train carriage</td>
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<td>0</td>
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<td>0</td>
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<tr>
<td>University campus—Eastside Bar</td>
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<td>5</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>3</td>
</tr>
<tr>
<td>University campus—RSM Workshop</td>
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<td>0</td>
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<td>0</td>
<td>1</td>
<td>0</td>
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<tr>
<td>Primary school</td>
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<tr>
<td>Total samples</td>
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<td>5</td>
<td>21</td>
<td>25</td>
<td>13</td>
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<tr>
<td>Total positive</td>
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<td>5</td>
<td>0</td>
<td>3</td>
<td>0</td>
<td>2</td>
<td>6</td>
</tr>
</tbody>
</table>

PM, particulate matter; TSP, total suspended particulate.
## Table 4  
PCR-positive SARS-CoV2 RNA samples

<table>
<thead>
<tr>
<th>Sample ID</th>
<th>Sampler type</th>
<th>Filter type</th>
<th>Site</th>
<th>Date of collection (lockdown measure)*</th>
<th>Seven-day average COVID-19 cases in area†</th>
<th>Volume air sampled (m³)</th>
<th>Copy no./Volume air (m³)</th>
<th>PM2.5 (µg/m³) (mean±SD)</th>
</tr>
</thead>
<tbody>
<tr>
<td>CV30</td>
<td>Biospot VIVAS Condensation</td>
<td>NP room on respiratory ward</td>
<td>25 March 2021 (step 1)</td>
<td>12.7</td>
<td>0.96</td>
<td>899</td>
<td>NA</td>
<td></td>
</tr>
<tr>
<td>CV40</td>
<td>SKC Button sampler</td>
<td>Gelatin</td>
<td>Respiratory ward</td>
<td>17 April 2021 (step 2)</td>
<td>9.9</td>
<td>0.12</td>
<td>64741</td>
<td>NA</td>
</tr>
<tr>
<td>CV46</td>
<td>Harvard impactor PM2.5 ≥0.1 &lt;2.5 µm</td>
<td>Medical day unit</td>
<td>12 May 2021 (step 2)</td>
<td>5</td>
<td>43.2</td>
<td>533</td>
<td>1.1±0.3</td>
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</tr>
<tr>
<td>CV47‡</td>
<td>Harvard impactor PM10 &gt;2.5 ≤10µm</td>
<td>Medical day unit</td>
<td>12 May 2021 (step 2)</td>
<td>5</td>
<td>43.2</td>
<td>23559</td>
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<tr>
<td>CV48‡</td>
<td>MVS Teflon 2.5</td>
<td>Medical day unit</td>
<td>12 May 2021 (step 2)</td>
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<td>7.2</td>
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<tr>
<td>CV54</td>
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<td>0.96</td>
<td>17174</td>
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<td>ICU</td>
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<td>7.2</td>
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<td>ICU</td>
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<td>CV67</td>
<td>MVS Teflon 2.5</td>
<td>ED waiting room</td>
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<td>151809</td>
<td>6.9±2.4</td>
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<td>ED waiting room</td>
<td>02 June 2021 (step 3)</td>
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<td>CV110</td>
<td>Button sampler</td>
<td>Teflon</td>
<td>Northern Line (Archway-Tottenham Court Road) 20 min</td>
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<td>140.9</td>
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<tr>
<td>CV112</td>
<td>Button sampler</td>
<td>Teflon</td>
<td>Piccadilly Line (Leicester Sq-Gloucester Rd) 20 min</td>
<td>02 August 2021 (step 3)</td>
<td>83.0</td>
<td>0.12</td>
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<tr>
<td>CV114</td>
<td>Button sampler</td>
<td>Teflon</td>
<td>Northern Line (Kennington-Leicester Square) 20 min</td>
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<td>171.7</td>
<td>0.12</td>
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<tr>
<td>CV116</td>
<td>Harvard impactor PM10 &gt;2.5 ≤10µm</td>
<td>Medical day unit</td>
<td>30 November 2021 (no restrictions)</td>
<td>107.6</td>
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<td>158</td>
<td>1.4±1.9</td>
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<td>Harvard impactor PM2.5 ≥0.1 &lt;2.5 µm</td>
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<td>30 November 2021 (no restrictions)</td>
<td>107.6</td>
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<td>30 November 2021 (no restrictions)</td>
<td>107.6</td>
<td>43.2</td>
<td>513</td>
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<tr>
<td>CV120</td>
<td>MVS Teflon</td>
<td>Medical day unit</td>
<td>30 November 2021 (no restrictions)</td>
<td>107.6</td>
<td>7.2</td>
<td>709</td>
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<tr>
<td>CV121</td>
<td>PDR (GMF) Microglass</td>
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<td>30 November 2021 (no restrictions)</td>
<td>107.6</td>
<td>2.88</td>
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<td>Button sampler</td>
<td>Teflon</td>
<td>Northern Line 30 min (Archway-Leicester’s Square)</td>
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<td>CV123</td>
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<td>Teflon</td>
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<td>0.12</td>
<td>143892</td>
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*UK government four-step roadmap for lifting restrictions and mapping a route back to normal life (https://www.gov.uk/government/publications/covid-19-response-spring-2021): step 1—travel prohibited, work from home, schools resume, limited university return, outdoors gatherings and sports permitted but limited to six people. Step 2—re-opening of non-essential retail, personal care premises (hairdressers/nail salons), public buildings (libraries, community centres), leisure facilities. Step 3—gatherings of up to 30 people permitted, re-opening of indoor entertainment venues, hotels, hostels, indoor events with capacity of 1000 people permitted. Step 4—gradual re-opening of nightclubs, eased restrictions on large events, outdoor meeting encouraged.
†Seven-day average number of COVID-19 cases were obtained from the coronavirus data dashboard (https://coronavirus.data.gov.uk/details/cases) developed by the UK Health Security Agency.
‡Samples CV47 and CV48 were successfully sequenced and found to be associated with the S-variant B1.1.7. Spike P681H detected was associated with a Nigerian mutation (N gene R203K, G204R, S235F, D288G).
ED, emergency department; ICU, intensive care unit; MVS, mini volume sampler; NA, not available; NP, negative pressure; PM, particulate matter.
The highest concentrations of virus were found on samples collected in the PM2.5 fraction (figure 3A,B). The second highest were found using TSP, with the next highest in the PM10 fraction. The lowest concentrations of virus were found in the PM1 fraction.

Performance of air samplers in detection of SARS-CoV-2
Out of 207 samples, most were collected using PM samplers (152 samples; 73% of total) compared with liquid TSP samplers (54 samples; 27% of total). PM samplers using PTFE and polyurethane foam filters more frequently detected SARS-CoV-2 (n=17) compared with liquid-based TSP samplers (n=3), with a detection rate of 11% vs 6% (figure 3C).

DISCUSSION
We detected SARS-CoV-2 RNA in air samples collected from several hospitals and public spaces in London during the partial lifting of restrictions, following the third national lockdown. Most of the positive samples were collected inside hospitals, first in an ICU with patients with COVID-19 receiving treatment and then in respiratory wards, as has been previously reported. In hospital areas, we also picked up the virus in non-COVID-19-associated waiting areas in clinics, in this case, in the waiting area of an oncology clinic. We found the presence of SARS-CoV2 on airborne PM picked up in the carriage but not on the platforms is of great interest because levels of PM on the London Underground are known to be very high. Previously, SARS-CoV2 virus have been detected from air samples collected between May and July 2020 inside buses and subway trains in Barcelona, Spain, particularly on PM2.5. We did not pick up virus from the other sites (university campus and primary school), possibly owing to policies in place which encourage regular testing for

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**Table 5** Maximum detectable SARS-CoV-2 concentrations

<table>
<thead>
<tr>
<th>Study</th>
<th>Maximum detectable SARS-CoV-2 concentration (RNA copies/m³)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Zhou et al⁷</td>
<td>219</td>
</tr>
<tr>
<td>Hu et al⁷</td>
<td>11 200</td>
</tr>
<tr>
<td>Lednicky et al¹³</td>
<td>0.74</td>
</tr>
<tr>
<td>Lednicky et al¹⁴</td>
<td>31 400</td>
</tr>
<tr>
<td>Chia et al¹⁸</td>
<td>3 380</td>
</tr>
<tr>
<td>Liu et al¹⁹</td>
<td>113</td>
</tr>
<tr>
<td>This study</td>
<td>7 072 284</td>
</tr>
</tbody>
</table>
detection of COVID-19 infection, hence a lower likelihood of infected persons detected earlier and isolation occurring sooner. Negative air samples for the presence of SARS-CoV2 in different indoor public places in Italy has also been reported.\(^{21}\) Moreover, we did not detect virus in the railway station, possibly owing to the better ventilation conditions due to more open space and lower particulate concentrations.

We used a wide range of air samplers and sampling techniques because there is no general agreement as to the best method of sampling for SARS-CoV2 detection in the air.\(^{22}\) The highest concentrations of SARS-CoV2 virus collected on 7 samples that was above 100000 copy numbers per m\(^3\) occasions may have arisen from our longer sampling periods, particularly when we used the size-fractionated PM samplers for up to 8 hours of collection. Surprisingly, we were able to detect the virus after only 30 min while using portable air samplers. We observed higher pick-up rates on size-segregated PM samplers compared with TSP collectors. When assessing size-segregated PM samplers, the highest pick-up rates were seen using the mini volume sampler (18% pick-up rate) and pDR-1500 (14% pick-rate), both of which collect samples at lower flow rates (2–5 L/min) with lower pick up using the Harvard impactor (6%, sample rate of 30 L/min), although a proportionately higher number of samples were collected by the Harvard impactor. As depicted in table 4, on the 2 days when virus was detected in both PM10 (containing PM \(>2.5 \leq 10\) µm) and PM2.5 (containing PM \(\geq 0.1 <2.5\) µm) fractions from the Harvard impactor as depicted in table 4, the amount of virus was higher in the PM10 fraction compared with the PM2.5 fraction as would be predicted. Interestingly, the next highest pick-up rates were noted in the portable SKC personal sampler (15% pick-up rate), with five out of the six positive samples collected on PTFE filters. Therefore, we were more likely to detect SARS-CoV2 when collecting at lower flow rates using techniques that impact PM and segregate them in terms of size. This higher pick-up rate may reflect the longer sampling periods when using particulate samplers, increasing the sampling volume and likelihood of capturing virus in the air. Alternatively, high flow rates may damage the virus.

Our detection of SARS-CoV-2 on PM, particularly PM2.5 and PM10 supports the notion of an interplay between virus and PM, suggesting there may be an interaction between SARS-CoV2 and PM.\(^{23}\) PM has been reported to interact with pathogens and may act as a vector for disease transmission.\(^{24-26}\) Most (75%) of our positive samples were collected onto filters which contained PM2.5 fine particles, in agreement with findings of Kayalar et al in Turkey.\(^{11}\) In the outdoor study performed in Bergamo Italy, samples collected were positive in PM10 samples for SARS-CoV2, because PM10 was the only particulate fraction that was collected.\(^{12}\) This link would provide support for the epidemiological studies in China and the USA that indicate that people living in high pollution areas particularly with high concentrations of PM2.5, experience

**Figure 3** SARS-CoV-2-positive air samples. Panel A demonstrates the mean concentration of virus detected on TSP and on each PM size fraction. Panel B demonstrates the concentration (copy/m\(^3\)) of SARS-CoV-2-positive samples collected using different samplers. Each point represents a single measurement, collected by the indicated instrument sampler. Panel C shows the % pick up of SARS-CoV-2-positive sample for TSP and SF PM samplers and for each individual air sampler. BS, button sampler; HI Harvard impactor; MVS, mini volume sampler; PM, particulate matter; PS, personal sampler; PTFE, polytetrafluoroethylene filter; SF, size fractionated; TSP, total suspended particulate.
more severe COVID-19 disease with higher mortality rates should they get infected with SARS-CoV2 virus.27 28 One possibility is that PM2.5 may act as a conduit for the virus to reach the small airways and the alveoli, thereby favouring the development of pneumonia.29 In addition, exposure to PM may upregulate expression of ACE-2, the receptor which the virus binds to via its spike protein,30 indicating that PM may also increase susceptibility to SARS-CoV2 infection. Thus, the COVID-19 pandemic highlights the need for lowering the levels of PM2.5, to limit the spread of SARS-CoV2 spread.31

Despite this co-existence of SARS-CoV2 and PM2.5, we did not culture SARS-CoV2 after inoculating to Vero E6 cells despite some of the high levels of SARS-CoV2 copies observed in many samples. A possible explanation could be a lack of viability of the suspended virus sampled. While the SARS-CoV2 virus may remain viable for up to 3 hours in aerosols generated into air,9 a recent study found that there was a rapid loss of infectivity of the aerosols within minutes due to the elevation of pH as the aerosol evaporates.32 However, stability of the virus in connection with PM is not known. It has been reported that PM2.5-derived reactive oxygen species resulting from interaction with epithelial cells may impair the structure and survival of influenza A that binds to PM2.5.33 It is possible that interactions with PM may alter the survivability of the virus and, therefore, the determination of the minimum concentration of SARS-CoV2 virus associated with PMs to propagate viral replication in Vero E6 cells or lung epithelial cells is needed.

The pick up of positive samples in these public/semi-public places is very much dependent on the presence of any infected asymptomatic or mildly symptomatic persons and producing aerosols containing SARS-CoV2 virus. One limitation of the study is our inability to determine whether there were any positive cases present or circulating within the spaces sampled. This may well explain why we did not detect any SARS-CoV2 in the bars, university campus rooms or in the school that we sampled. In addition to the source of the virus, other factors that will determine the amount and survivability of the virus are the environmental conditions such as temperature, humidity and airflow.34 It is unclear as to whether large airborne titres of virus are being produced by multiple infected people residing in the same space, or from potential ‘super-spreaders’. Moreover, knowing whether an infected case was present, would help us estimate how long respirated virus remains airborne.

Another limitation is the use of a single primer targeting the N-gene of SARS-CoV2 in processed airborne samples, via RT-qPCR. There are data to suggest primers targeting the N-gene is highly sensitive, with increased risk of false positives.35 This increases the risk of overestimating the viral copy number. On the other hand, inability to extract 100% of particles from all filter types could mean that virus concentrations have been underestimated, as we were unable to extract the entire sample. In two samples collected, we were able to confirm, by full sequencing, that the variants collected were the dominant variants at that time, that is, the S-variant B.1.1.7, that confirm that SARS-CoV2 virus was detected.

In summary, we detected SARS-CoV2 RNA virus in size-fractionated PM samples, particularly in the fine fractions of PM collected from hospital waiting areas and wards and in London Underground train carriage. This indicates that SARS-CoV2 can circulate in the air, but whether it is active needs further work such as elucidation any potential interactions of PM2.5 with SARS-CoV2 in the air and at the lung epithelial surface. Air sampling, using size-fractionated PM samplers or portable air samplers, may be important in determining the transmission potential of SARS-CoV2. In addition, as a matter of precaution, the wearing of face masks during such periods would be recommended, particularly in indoor and semi-open environments.

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Contributors. KFC, CP and PK conceived the idea; KFC obtained the funding; IA, FF, CP, AP and KFC discussed the approach to data analysis; HA-W and GK collected the data; HA-W, GK, PKB and PK analysed the data; PKB and RW performed RT-qPCR; FH, ER and VS performed the virus culture; KFC, HA-W and GK wrote the manuscript; all authors contributed to its finalisation and agreed with the final version. All authors gave final approval of the manuscript, had full access to all the data in the study and had final responsibility for the decision to submit for publication. The corresponding author acts as a guarantor. The corresponding author attests that all listed authors meet authorship criteria and that no others meeting the criteria have been omitted. The corresponding author (the manuscript’s guarantor) affirms that the manuscript is an honest, accurate and transparent account of the study being reported; that no important aspects of the study have been omitted and that any discrepancies from the study as planned (and, if relevant, registered) have been explained.

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REFERENCES

SARS-CoV2 in public spaces in West London UK during COVID-19 pandemic

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Supplemental materials
Methods

Liquid-based Total Suspended Particulate (TSP) Samplers

Unsegregated total suspended particulates (TSP) were collected using the commercially-available liquid-based samplers including the Biospot-VIVAS (Aerosol Devices Inc., Ft. Collins, CO), the Sartorius MD8 (Sartorius AG, Germany) and the Coriolis μ air sampler (Bertin Technologies). The Biospot VIVAS has been shown to be efficacious in collecting viral particles [11] and airborne particles using a water vapour condensation method, at a rate of 8 litres per minute (L min⁻¹). Air initially passes through a cool temperature conditioner, followed by passage through the initiator at 30°C which coalesces particles as small as 8 nm into larger droplets greater than 2 µm in diameter. The enlarged particles are then collected through a set of nozzles onto 1.5 mL of liquid collection media. The collection media used, composed of 1.5mL 1x phosphate buffered saline (PBS), 0.5% (w/v) bovine albumin fraction V and sucrose added to a final concentration of 0.2M, has been used to successfully culture SARS-CoV-2 [1]. Liquid samples were stored at 4°C and transported on ice for viral RNA extraction and SARS-CoV-2 PCR testing.

The Sartorius MD8 (Sartorius AG, Germany) sampled air at a rate of 30 L min⁻¹ for 30 minutes, impacting particles onto a sterile gelatin filter (80 mm diameter, 3 µm pore size, type 80-ACD, Sartorius AG). After sampling, the gelatin filter was transferred aseptically into a Petri dish and transported on ice and stored in a refrigerator at 4°C for later processing.

We also used the Coriolis μ air sampler (Bertin Technologies) which collects air at 100 L min⁻¹. Samples were collected every 30 minutes into a conical vial containing 15 mL phosphate-buffered saline. Samples were stored at 4°C with viral RNA extraction fluid and qRT-PCR performed on neat samples.
Size-segregated Particulate Matter Samplers

Size-segregated particles were collected using a MiniVOL sampler, (Airmetrics, Springfield, OR, USA) Harvard Cascade Impactor (HCCI) and personal DataRAM (pDR-1500). The MiniVOL sampler (MVS) (Airmetrics, Springfield, OR, USA) was equipped with PM$_{10}$ and PM$_{2.5}$ impactors and collects PM$_{2.5}$ particles at a flow rate of 5 L min$^{-1}$ using a double-diaphragm pump with laminar flow valve technology. This means that the PM fraction collected will be $\leq 2.5 \mu$m. The particles were collected onto a 47 mm polytetrafluoroethylene (PTFE) filter (TISCH Scientific, 201 S Miami Ave, Cleves, OH 45002, USA). Particles were collected on Teflon filters because they are hydrophobic in nature, chemically resistant, have high initial particle capture efficiency across different flow rates and are suitable for gravimetric, chemical and microscopic analysis of PM.

The Harvard Impactor collected size-segregated particles of PM$_{10}$ (coarse), PM$_{2.5}$ (fine) and PM$_{1}$ (ultrafine) on polyurethane foams (PUF) (Merry weather Foam, OH, USA) at a rate of 30 L min$^{-1}$ with particles fractionated at three different impaction stages. Particles $\leq$100 nm (PM0.1) were collected simultaneously onto 47 mm PTFE filters (TISCH Scientific 201 S Miami Ave, Cleves, OH 45002, USA) at the final stage of this cascade impactor. This means that the coarse fraction collected will contain PM $>2.5\leq10 \mu$m, the fine fraction PM $\geq0.1<2.5 \mu$m, and the ultrafine particles below 100nm of PM$<0.1 \mu$m.

The personal DataRAM™ (pDR-1500) aerosol monitor (Thermo Scientific, Franklin, MA, USA) is a sensitive nephelometric monitor with a cyclone inlet for measurement and collection of PM$_{2.5}$ particles. The pDR-1500 reported the average PM$_{2.5}$ concentration every 1 min and collected particles onto 37 mm glass fibre filters.
through the sensing zone at a flow rate of 1.52 L min$^{-1}$ for a cut size of 2.5 µm diameter. All the filters used in the above-mentioned size segregated particle samplers were allowed to equilibrate in a weighing room with controlled temperature (21°C) and relative humidity (30–40%) for a minimum of 24 hr prior to weighing before and after sampling. After each sampling session, filters were removed using forceps and inserted into sterile Petri dishes and sealed using parafilm. These were transported on ice to a level 2 laboratory for particle extraction and SARS-CoV-2 PCR testing.

**Portable Samplers**

We also sampled using the portable SKC Button sampler (SKC LTD, Dorset, UK). Air was extracted at a rate of 4 L min$^{-1}$ using the SKC Airchek touch pump, which was attached to the outlet of the button or a filter cassette, loaded with a 37mm PTFE filter. The sampler was attached to an individual’s clothing, on their chest, ~2 cm under their clavicle, whilst wearing an FFP3 mask. Devices were calibrated using an adaptor by attaching the Button sampler to the inlet of the button, and the inlet of the calibration adaptor to a HVAC system prior to use. Air samples were collected directly onto a dissolvable 25 mm gelatin filter (1.0µm pore size, SKC LTD, Dorset) when sampling with the button, or unto the 37mm PTFE filter (0.3µm pore size, SKC LTD, Dorset) pre-loaded to a filter cassette, using an appropriate adaptor as per manufacturer’s instructions. The button sampler collects ‘inhalable’ particles with a 1µm-100µm aerodynamic diameter as defined by the British Standards Institution (BS EN 481:1993). The PTFE loaded cassette collects total suspended particulate with an aerodynamic diameter above 0.3µm.

**Post-sampling processing**
**PUF and TF filters**

Filters were removed from air samplers using forceps and placed in a Petri dish at the sampling site. PBS was added to the filters in a class II biological safety cabinet (10 ml to polyurethane foam (PUF), 5 ml to PFTE) and sealed with parafilm. This was then placed on a shaker for 30 minutes at 70 RPM to gently transfer particles from the filter into the solution. After 30 minutes, the solution was transferred into a 10 ml Falcon tube in an L2 HEPA-filtered hood using a pipette. 1 ml of the solution was immediately sent for culture on Vero-E6 cells in a 1.5 ml Eppendorf, with the remaining solution stored at -80ºC for RNA extraction and RT-qPCR.

**Gelatin filters**

80 mm gelatin filters were placed into a Petri dish with 10 ml of PBS added whereas 37 mm gelatin filters from portable samplers were placed in 50 ml falcon tubes with 5 ml of PBS added. They were then placed in a shaker at 70 RPM and warmed to 28ºC for 30 minutes to dissolve the filter. The remaining solution was transferred into a 10 ml Falcon tube in a L2 HEPA-filtered hood using a pipette. 1 ml of the solution was immediately sent for culture on Vero-E6 cells in a 1.5 ml Eppendorf tube, with the remaining solution stored at -80 ºC for RNA extraction and RT-qPCR.

**Biospot VIVAS and Coriolis**

Air particles from the Biospot VIVAS were directly collected in liquid medium in a small petri dish and conical flask respectively. After sampling, an equal volume of the solution (between 500-1000 μl) was transferred into three separate 1.5ml Eppendorf tubes. One Eppendorf tube containing solution was immediately sent for culture on Vero-E6 cells, with the remaining stored at -80ºC for RNA extraction and RT-qPCR.
Reference: