Vimentin as a target for the treatment of COVID-19

Zhenlin Li, Denise Paulin, Patrick Lacolley, Dario Coletti, Onnik Agbulut

Abstract
We and others propose vimentin as a possible cellular target for the treatment of COVID-19. This innovative idea is so recent that it requires further attention and debate. The significant role played by vimentin in virus-induced infection however is well established: (1) vimentin has been reported as a co-receptor and/or attachment site for SARS-CoV; (2) vimentin is involved in viral replication in cells; (3) vimentin plays a fundamental role in both the viral infection and the consequent explosive immune-inflammatory response; and (4) a lower vimentin expression is associated with the inhibition of epithelial to mesenchymal transition and fibrosis. Moreover, the absence of vimentin in mice makes them resistant to lung injury. Since vimentin has a twofold role in the disease, not only being involved in the viral infection but also in the associated life-threatening lung inflammation, the use of vimentin-targeted drugs may offer a synergistic advantage as compared with other treatments not targeting vimentin. Consequently, we speculate here that drugs which decrease the expression of vimentin can be used for the treatment of patients with COVID-19 and advise that several Food and Drug Administration-approved drugs be immediately tested in clinical trials against SARS-CoV-2, thus broadening therapeutic options for this type of viral infection.

Introduction
The COVID-19 has triggered a global public health crisis with more than 10 million infected people up to date (30 June 2020) worldwide and a global incidence rate which is still growing. SARS-CoV-2 infection causes an explosive immune response, characterised by a cytokine storm, a subsequent progression to lung injury and an acute respiratory distress syndrome leading to death. Despite many promising therapeutic options for the treatment of COVID-19, anti-SARS-CoV-2 drugs or vaccines are still under investigation. Vimentin, known as a cytoskeletal protein belonging to the intermediate filament (IF) family, plays an important role in stabilising intracellular architecture through its mechanical role in cell plasticity and organelle anchoring, in the metabolism of lipids and in cell remodelling through its interaction with signalling molecules and components of gene regulatory networks. At cellular level, vimentin has been shown to be implicated in cell proliferation, stiffness, adhesion, migration, differentiation, senescence and apoptosis. Vimentin also plays a role in inflammation and immune responses, as well as in the epithelial to mesenchymal transition (EMT); the latter, in turn, being involved in the opening of epithelial barriers and cell migration. Strikingly, intracellular vimentin has been shown to be implicated in the processes of virus fusion, replication and assembly (for a review, see Ramos et al). Vimentin was additionally found to be located outside of the cell, where it serves as an attachment site for viral proteins, in the majority of cases favouring viral binding/entry. One possible exception was reported for the human papillomavirus type 16 (HPV-16) infection, in which vimentin actually had the opposite effect. In this case, the authors reported that cell surface vimentin interfered with viral binding to its receptor, thus having a protective effect against viral infection. However, the role of vimentin for this specific virus is debated and contradictory reports have been published. As evidence to the contrary, Jacobs et al experimentally reduced the level of vimentin on the surface of cervical cancer cells by exposing the cells to the hookworm Nippostrongylus brasiliensis, with the result of decreasing the internalisation of the HPV-16 in these cells. One of the proposed mechanisms underlying this observation was the production of T helper-derived cytokines (eg, interleukin (IL)-4) associated with the helminth infection, which could, in turn, have modified the expression of HPV receptors in cancer cells. During the submission period of the present manuscript in which we propose vimentin as a potential therapeutic target against viral infection, another two papers were written advocating the same hypothesis. Vimentin was also been previously proposed as a potential therapeutic target in cancer treatment and other disease conditions such as fibrosis. The upregulation of vimentin typically observed in oncogenic transformation and...
its involvement in cancer cell migration highlighted the possibility that vimentin is not only a marker but also a player in the development of cancer (reviewed in Refs 10 and 19). Therefore, molecules such as withaferin A, FvE1 and simvastatin were used to perturb the expression and/or the assembly of vimentin filament and led to the downregulation of soluble vimentin, accompanied by decreased fibrosis, the disruption of mitotic cells, filament disorganisation and cell death.20–22

This paper summarises the scientific evidence indicating that reducing the expression of vimentin could be a powerful therapeutic option for COVID-19, since it could hamper SARS-CoV-2 infection and, at the same time, decrease the immunological response that is ultimately responsible for the often lethal acute respiratory distress syndrome. This hypothesis can be readily tested through clinical trials since Food and Drug Administration (FDA)-approved pharmacological treatments targeting vimentin for other pathologies are already available.

Vimentin is a co-receptor for SARS-CoV and possibly for SARS-CoV-2

ACE 2 (ACE2) is a cellular receptor for the SARS-CoV spike protein.23 However, it has been shown that ACE2 is not sufficient to make host cells susceptible to infection24–26 and vimentin has been proposed as a co-receptor for the entry of SARS-CoV into cells.27 Indeed, the authors reported that the SARS-CoV virus enhances cell surface vimentin expression and, importantly, showed the existence of a direct interaction between vimentin and the SARS-CoV spike protein during viral entry.13 Moreover, SARS-CoV and SARS-CoV-2 share similar spike protein sequences and the same cell entry route.27 Interestingly, it has been recently reported that two proteins that are well known to be involved in the entry of SARS-CoV-2 into the cell—ACE2 and TMPRSS2—are both expressed by two cell types, that is, lung type II pneumocytes and nasal goblet secretory cells28; it is worth noting these two cell types also express high levels of vimentin.29–30 In addition, a potential interaction between vimentin and the SARS-CoV-2 spike protein is revealed in a SARS-CoV-2 protein–protein interaction map (see supplemental data).31

Vimentin starts being expressed in the early stages of embryonic development by highly plastic precursor cells, whereas in postnatal life it is found in fibroblasts, endothelial cells, smooth muscle cells and in the lining epithelial cells of the lung, gut and other mucosa.29 30 32 33 Here vimentin expression is quickly increased in response to viral infection and inflammatory stimuli34–36 since vimentin belongs to the ‘immediate early genes’ family, a group of genes which are quickly activated in the presence of infection and inflammation. In this condition, due to its translocation to the cell surface, vimentin works as a co-receptor or attachment site for several viruses, including the Japanese encephalitis virus,37 the porcine reproductive and respiratory syndrome virus,38 the enterovirus 71,39 cowpox mosaic virus,40 dengue virus41 as well as the coronavirus.13 Interestingly, an upsurge of vimentin expression accompanies, and favours, viral entry of the majority of viruses including SARS-CoV13 with the exception of HPV-16 in which viral entry is actually hampered by an increase in vimentin expression.17 Consistently, interfering with vimentin expression or treating cells with the neutralising ‘anti-vimentin’ antibodies attenuates some types of viral infection.38 39 18 37 38 However, it is unknown if vimentin is involved in the infection of the Middle East respiratory syndrome coronavirus (MERS-CoV) which uses dipeptidyl peptidase 4 as the host cell receptor.40

Vimentin is involved in different steps of viral replication or assembly since it colocalises with viral capsid proteins in the endoplasmic reticulum of the infected cells,41 cooperating to produce virus particles.42 It is reported that vimentin is implicated—through the control of lysosomal trafficking and reduced translocation of viral ribonucleoproteins—in the progression of the influenza A virus, acting as a restriction factor for viral replication.43 44 In addition, vimentin influences the hepatitis C virus (HCV) and Thelmer’s virus replication through its interaction with virus core proteins.45 46 A strong reduction of HIV-1 replication was reported in vimentin knockdown cells, showing that vimentin is necessary for viral replication.47 Vimentin contributes to EMT, a process by which epithelial cells, including those lining the lung mucosa, lose their polarity and adhesion. EMT thus confers migratory and invasive properties to the cells in a variety of pathological conditions such as viral infections, angiogenesis, chronic autoimmune inflammatory diseases, inflammatory bowel diseases, chronic obstructive pulmonary disease, cystic fibrosis and cancer. Overall, EMT promotes the creation of a suitable environment for viral infection and inflammation.

Vimentin expression is regulated by a plethora of transcription factors such as NF-kB (nuclear-factor-kappa B), Stat3 (signal transducer and activator of transcription 3), ZBP-89 (Kruppel-type zinc-finger family transcription factor), Smads (smallmothers against decapentaplegic transcription factors), AP-1 (activator protein 1), Sp1 (proximal specificity protein 1) and PEA3 (polyomaenhancer activator 3)—see figure 1. In response to viral infection or other insults, vimentin expression quickly increases. There are other transcription factors, such as Snail, Twist, ZEB2 and Slug,48–52 that regulate vimentin expression during EMT so that the overexpression of vimentin and N-cadherin in epithelial cells leads to the loss of the epithelial phenotype.

In addition, vimentin expression is also regulated epigenetically through the chromatin modifications of its promoter.53 These modifications have been studied in cancer cells since they promote EMT—a crucial event in the formation of tumour metastases. In particular, the methylation level of the vimentin promoter inversely correlates with the expression of vimentin in gastric...
cancer. Finally, vimentin expression is also regulated by post-transcriptional mechanisms including that of non-coding microRNAs.

Vimentin actively participates in inflammation and immune response

Targeting vimentin could have additional effects against the progression of COVID-19 by directly modulating the inflammatory response of patients with COVID-19. Indeed, a subgroup of these patients suffer from a cytokine storm syndrome, that is, an excessive cytokine production sustained by a positive feedback loop producing an inflammatory response, often leading to respiratory failure from acute respiratory distress syndrome.

The NLRP3 inflammasome (nucleotide-binding and oligomerisation domain-like receptor protein 3 inflammasome) is an intracellular multiprotein complex that triggers the immune response of the host against pathogens including viruses and is implicated in acute lung injury. It has been demonstrated that vimentin is a key regulator of the NLRP3 inflammasome by direct interaction. Important events in lung inflammation and the onset of injury are the increase in IL-1β levels, the permeability of the endothelial–alveolar epithelium barrier and irreversible fibrosis. The relevance of vimentin in these responses was demonstrated in animal models in which the inflammatory response of vimentin-knockout mice to lipopolysaccharide (LPS), bleomycin or asbestos treatment was considerably lower, entailing a less significant lung injury. Leucocyte adhesion to vascular endothelium and platelets is an early step in acute inflammatory response and attenuation of this adhesion may be beneficial in acute lung injury. By increasing circulating IL-1β—the major proinflammatory cytokine—vimentin is responsible for activating and recruiting inflammatory cells. In this context, it was shown that the vimentin IFs of both endothelial cells and lymphocytes form an anchoring structure between these two cell types. Further evidence of the importance of vimentin for leucocyte extravasation comes from the report that the transendothelial migration of blood T and B lymphocytes is markedly reduced in vimentin-deficient lymphocytes and endothelial cells. On the other hand, the presence of vimentin on the platelet and endothelial cell surface also serves as an adhesive receptor for the von Willebrand factor (VWF) and causes the binding of platelets to the subendothelial collagen and the subsequent intravascular generation of thrombin. The latter being responsible...
for microthrombus formation and contributing to a non-
haemostatic effect, that is, the inflammatory response.

The inhibition of the vimentin–VWF interaction,
by anti-vimentin antibodies, is effective in interfering
with the VWF-mediated platelet adhesion to different
matrices.57 58 The administration of exogenous recombi-
nant human vimentin, binding specifically to P-selectin,
stops leucocyte adhesion to platelets and endothelium
by blocking P-selectin interaction with P-selectin glyco-
protein ligand, ultimately decreasing endotoxin-induced
acute lung injury.59 The oxidised form of membrane-
bound vimentin is a marker of senescent cells,60 which,
as such, produce a plethora of potentially harmful proin-
flammatory cytokines, chemokines and growth factors.
Virus incubation times in senescent cells is longer, as
observed in the case of HPV-1661 and studying corona-
virus infection in aged cells has been proposed, since this
important issue for this virus is still unexplored.9

As shown above, vimentin is expressed by many cell
types, it has multiple cellular localisations, it plays multiple
roles in cell behaviour by interacting with a plethora of
molecular partners. The complexity of the functions
exercised by vimentin, especially as concerns viral infec-
tion, is exemplified by contradictory reports in the lit-
erature showing opposite roles for vimentin, depending
on the type of virus and the specific partners involved in
its direct or indirect interactions. In the following para-
graph and in table 1, we report and comment on a list
of vimentin-targeting drugs, which are globally known to
downregulate vimentin expression, since we think that
such a downregulation would be beneficial. However,
we decided to provide only a few significant case by case
comments. Each single drug, depending on the type of
virus and host cell type concerned, should be further
investigated.

**Drugs targeting vimentin could combat coronavirus-related
effects**

Based on the fact that vimentin is important for viral
infection and the associated inflammatory response,
we have searched the literature for compounds targeting
vimentin so as to select drugs that may treat COVID-19.
Some of these drugs are as common and as easily avail-
able as melatonin, which makes our hypothesis imme-
diately verifiable through clinical trials on COVID-19
specifically.

Some of the 49 compounds that we have identified
(table 1) are completely new molecules that have recently
started to be studied, others have never been employed
in clinical practice although they are referenced by
the FDA, while others are already being tested in clin-
ical trials. However, all these molecules in table 1 affect
vimentin levels, as demonstrated by western blot and/or
real-time PCR analysis. Several of these compounds have
been identified and characterised in in vitro experiments
only, while others have been validated in vivo against cell
growth and dispersion in cancer, and for their capacity to
inhibit EMT which, as mentioned, amounts to fighting
inflammation.

Interestingly, a significant fraction (16 out of 49) of
the drugs listed in table 1 have been reported to play a
role in acute lung injury and are briefly discussed below
for this reason. **Melatonin** markedly reduces pulmonary
injury and decreases the infiltration of macrophages
and neutrophils into the lungs of LPS-treated mice, by
inhibiting the NLRP3 inflammasome, in a model of acute
lung injury.62 Worth noting, it has already been specu-
lated that melatonin would be beneficial in patients with
COVID-19.63 **Niclosamide**, a cheap anthelmintic drug,
is effective against several viruses such as SARS-CoV,
MERS-CoV, Zika virus, HCV and human adenovirus.64 In
particular, niclosamide has been shown to be an inhibitor
of SARS-CoV 3CL protease, involved in viral replication.64

Endogenous hydrogen sulfide participates in the regulation
of important biological processes in the respiratory tract,
such as airway tone, pulmonary circulation, cell prolifer-
ation and apoptosis, fibrosis, oxidative stress and inflam-
mation.65 Since hydrogen sulfide exerts a broad-spectrum
antiviral activity, as well as having an anti-inflammatory
action, the controlled release of hydrogen sulfide from
donor compounds has been proposed as a treatment in
lung diseases.65 **Simvastatin** has protective vascular effects
in the lungs since it improves the function of the endo-
thelial barrier. The efficacy of simvastatin against LPS-
induced lung injury involves the stabilisation of the cell
cytoskeleton and adherent junctions.66 The ability of tri-
tolide to inhibit the proinflammatory NF-κB signalling
pathway makes it another promising therapeutic agent
for acute lung injury.67 **Sinomenine** attenuates septic shock-
dependent acute lung injury, probably thanks to the inhi-
bition of inflammation and oxidative stress.68 **Tanshinone
HA** inhibits the production of proinflammatory factors in
LPS-induced acute lung injury through the regulation of
calcium in pulmonary interstitial macrophages.69 **Ginse-
inoside Rg3** attenuates LPS-induced acute lung injury by
decreasing the production of proinflammatory factors and
increasing the synthesis of anti-inflammatory cyto-
kines, through the activation of the PI3K/AKT/mTOR
pathway downstream of the Mer receptor.70 **Icarin**
 reduces acute lung injury by enhancing the expression of
the glucocorticoid receptor alpha in lung tissues and
by inhibiting the expression of p65, c-Jun, Stat3, IL-6 and
tumour necrosis factor-alpha.71 The therapeutic window
of Valproic acid is narrower but interesting, since it is
effective in a mouse model of Gram-negative bacteria-
induced pneumonia.72 **Apigenin C-glycosides** inhibit acute
inflammation and apoptosis by suppressing the activa-
tion of the TLR4/TRPC6 signalling pathway in a murine
model of acute lung injury.73 **Inositol derivatives** present
in surfactant preparations diminish the activation of key
inflammatory pathways in lung diseases.74 **Pterostilbene
4-β-glucoside**, the glycosylated form of the antioxidant
pterostilbene, diminishes intracellular and mitochondrial
reactive oxygen species production, thus reducing
the inflammatory response to LPS.75 **Osthole** exerts
<table>
<thead>
<tr>
<th>Name of molecule</th>
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<th>Effects</th>
<th>Modes of action</th>
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<th>Reference</th>
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<tbody>
<tr>
<td>17-DMCHAG</td>
<td>Geldanamycin derivative</td>
<td>Antimigratory; antiproliferative</td>
<td>Inhibits HSP90; induces the proteasome-dependent degradation of androgen receptor</td>
<td>Human prostate cancer cells; tumour-bearing mice</td>
<td>79</td>
</tr>
<tr>
<td>Alpha-lipoic acid*</td>
<td>Acid; antioxidant</td>
<td>Inhibitor of EMT; antiproliferative</td>
<td>Inhibits TGF-β; activates AMPK and downregulates mTOR-S6 signalling</td>
<td>Human thyroid cancer cells; tumour-bearing mice</td>
<td>80</td>
</tr>
<tr>
<td>Apigenin†</td>
<td>Flavonoid derivative</td>
<td>Antiapoposis; pro-proliferative; antifibrotic</td>
<td>Decreases mIR-34a</td>
<td>Mouse mesothelial peritoneal cells</td>
<td>81</td>
</tr>
<tr>
<td>Berberine*</td>
<td>Alkaloid derivative</td>
<td>Inhibitor of EMT; antiproliferative</td>
<td>Decreases PI3/AKT, Ras-Raf-ERK and TGF-β1 signalling</td>
<td>Mouse neuroblastoma cells</td>
<td>82</td>
</tr>
<tr>
<td>Bergamottin</td>
<td>Furanocoumarin derivative</td>
<td>Inhibitor of EMT; antimigratory</td>
<td>Decreases PI3K/AKT/mTOR, TGF-β1 signalling</td>
<td>Human lung adenocarcinoma cells</td>
<td>83</td>
</tr>
<tr>
<td>Beta-asarone</td>
<td>Phenylpropanoid derivative</td>
<td>Inhibitor of EMT; antimigratory</td>
<td>Decreases hnRNP A2/B1, MMP-9, p-STAT3 expression</td>
<td>Human glioma cells</td>
<td>84</td>
</tr>
<tr>
<td>Beta-lapachone</td>
<td>Quinone derivative</td>
<td>Inhibitor of EMT; antimigratory; proapoptosis</td>
<td>Induces apoptosis through caspase-3, -8 and -9 activation and poly(ADP-ribose) cleavage; decreases MMP-2 and MMP-9</td>
<td>Mouse colon cancer cells; tumour-bearing mice</td>
<td>85</td>
</tr>
<tr>
<td>BHX</td>
<td>Pyrazoline derivative</td>
<td>Inhibitor of EMT; antimigratory; proapoptosis</td>
<td>Inhibits Wnt/β-catenin signalling</td>
<td>Human breast cancer cells; tumour-bearing mice</td>
<td>86</td>
</tr>
<tr>
<td>BMS345541</td>
<td>Selective I kappa B kinase inhibitor</td>
<td>Inhibitor of EMT; anti-inflammatory</td>
<td>Inhibits NF-κB/RelA pathway</td>
<td>Human epithelial cells; polyinosinic–polycytidylic acid-treated cells</td>
<td>87</td>
</tr>
<tr>
<td>Chrysotobibenzyl</td>
<td></td>
<td>Inhibitor of EMT; antimigratory</td>
<td>Reduces MMP-2 activity; induces zona occludens protein-1 and occludin expression</td>
<td>Human renal epithelial cells; db/db mice</td>
<td>88</td>
</tr>
<tr>
<td>Chrysin*</td>
<td>Flavonoid derivative</td>
<td>Inhibitor of EMT; antimigratory</td>
<td>Decreases integrins (β1, β3, αν), p-FAK, p-AKT, caveolin-1 expression</td>
<td>Human lung cancer cells</td>
<td>89</td>
</tr>
<tr>
<td>Cisplatin</td>
<td>Anticancer</td>
<td>Inhibitor of EMT; antimigratory; antiproliferative</td>
<td>Reduces YAP activity</td>
<td>Human colon cancer cells; tumour-bearing mice</td>
<td>90</td>
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<tr>
<td>Compound 11</td>
<td>Selective class I HDAC inhibitor</td>
<td>Inhibitor of EMT; antimigratory</td>
<td>Decreases p-AKT, p-ERK activity</td>
<td>Human colorectal cancer cells; tumour-bearing mice</td>
<td>91</td>
</tr>
<tr>
<td>Curcumin*</td>
<td>Polyphenolic compound</td>
<td>Inhibitor of EMT; antimigratory; antiproliferative</td>
<td>Decreases TGF-β3/Smad2/3 signalling</td>
<td>Human thyroid cancer cells</td>
<td>92</td>
</tr>
<tr>
<td>Cypripedin</td>
<td>Phenanthrenequinone derivative</td>
<td>Inhibitor of EMT; antimigratory</td>
<td>Decreases AKT/GSK-3β signalling</td>
<td>Human lung cancer cells</td>
<td>93</td>
</tr>
<tr>
<td>D-4F</td>
<td>Apolipoprotein A-I mimetic</td>
<td>Inhibitor of EMT; antifibrotic</td>
<td>Inhibits TGF-β1</td>
<td>Human alveolar epithelial cells</td>
<td>94</td>
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<tr>
<td>Dehydroepiandrosterone</td>
<td>Steroid</td>
<td>Inhibitor of EMT; antimigratory</td>
<td>Decreases N-cadherin and Snail expression</td>
<td>Human breast cancer cells; tumour-bearing mice</td>
<td>95</td>
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<tr>
<td>Dictamine</td>
<td>Alkaloid</td>
<td>Inhibitor of EMT; antimigratory; antiproliferative; proapoptosis</td>
<td>Decreases mTOR/p70S6K/elf4e and MAPK signalling</td>
<td>Human lung cancer cells</td>
<td>96</td>
</tr>
<tr>
<td>Ginsenoside Rg3†</td>
<td>Steroidal saponin</td>
<td>Inhibitor of EMT; antimigratory; proapoptosis</td>
<td>Inhibits LncRNA colon cancer-associated transcript 1 expression and PI3K/AKT signalling; inhibits MAPK and NF-κB signalling</td>
<td>Human colorectal cancer cells; lung cancer cells; tumour-bearing mice</td>
<td>97</td>
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<tr>
<td>Hydrogen sulfide†</td>
<td>–</td>
<td>Inhibitor of EndMT; decrease ER stress; vasodilator; cardioprotective</td>
<td>Decreases Smad2 and Src signalling</td>
<td>Human umbilical vein endothelial cells</td>
<td>99</td>
</tr>
<tr>
<td>Name of molecule</td>
<td>Type</td>
<td>Effects</td>
<td>Modes of action</td>
<td>Models</td>
<td>Reference</td>
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<tr>
<td><strong>Hydroxygenkwanin</strong></td>
<td>Flavonoid derivative</td>
<td>Antimigratory; antiproliferative</td>
<td>Activates p21 signalling</td>
<td>Human oral squamous cell carcinoma</td>
<td>44</td>
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<tr>
<td><strong>Icarin</strong>†</td>
<td>Flavonoid derivative</td>
<td>Antimigratory; antiproliferative; proapoptosis</td>
<td>Inhibits PI3K, AKT and MEK/ERK signalling; decreases miR-625-3p and MMP-9 expression</td>
<td>Human thyroid cancer cells</td>
<td>100</td>
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<tr>
<td><strong>Icariside II</strong></td>
<td>Flavonol glycoside</td>
<td>Inhibitor of EMT; antimigratory</td>
<td>Inhibits NF-κB and AKT/GSK-3β signalling</td>
<td>Human lung cancer cells; tumour-bearing mice</td>
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<tr>
<td><strong>Inositol</strong>†</td>
<td>Lipid</td>
<td>Inhibitor of EMT; antimigratory</td>
<td>Inhibits PI3K/AKT and NF-κB signalling</td>
<td>Human breast cancer cells</td>
<td>102</td>
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<tr>
<td><strong>Melatonin</strong>†</td>
<td>Hormone</td>
<td>Inhibitor of EMT; antimigratory</td>
<td>Inhibits PI3K/AKT signalling; decreases MMP-2, MMP-9, NF-κB p65 expression</td>
<td>Human ovarian cancer cells; gastric cancer cells; tumour-bearing mice</td>
<td>103 104</td>
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<tr>
<td><strong>Metapristone</strong>*</td>
<td>RU486 derivative</td>
<td>Antimigratory; antiproliferative; proapoptosis</td>
<td>Decreases AKT and ERK phosphorylation and Bcl-2; upregulates total p53 and Bax expression</td>
<td>Mouse skin melanoma cells</td>
<td>105</td>
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<tr>
<td><strong>Metformin</strong>†</td>
<td>Biguanide derivative; anti-diabetic</td>
<td>Inhibitor of EMT; antimigratory</td>
<td>Downregulates HIF-1alpha, CAIX, miR-34a, SNAIL1 and ZEB1; upregulates miR-200a, miR-200c and miR-429</td>
<td>Human cervical squamous cancer cells; human colorectal cancer cell</td>
<td>106 107</td>
</tr>
<tr>
<td><strong>Moscatilin</strong></td>
<td>Bibenzyl derivative</td>
<td>Inhibitor of EMT; proapoptosis</td>
<td>Decreases ERK and AKT activity; downregulates caveolin-1 level</td>
<td>Human lung cancer cells</td>
<td>108</td>
</tr>
<tr>
<td><strong>N-acetylcysteine</strong>*</td>
<td>Antioxidant</td>
<td>Reduce fibrosis; anti-inflammatory</td>
<td>Enhances antioxidant enzyme activities; downregulates oxidising enzymes’ expression</td>
<td>Crystalline silica-induced pulmonary fibrosis mice model</td>
<td>109</td>
</tr>
<tr>
<td><strong>Niclosamide</strong>†</td>
<td>Antihelmintic</td>
<td>Inhibitor of EMT; antimigratory</td>
<td>Inhibits Wnt/b-catenin signalling</td>
<td>Human oral squamous cell carcinoma</td>
<td>110</td>
</tr>
<tr>
<td><strong>Norcantharidin</strong></td>
<td>Cantharidin derivative</td>
<td>Inhibitor of EMT; antimigratory</td>
<td>Inhibits αvβ6-ERK-Ets1 signalling</td>
<td>Human colon cancer cells</td>
<td>111</td>
</tr>
<tr>
<td><strong>Osthole</strong>†</td>
<td>Coumarin derivative</td>
<td>Inhibitor of EMT; antimigratory; antiproliferative; proapoptosis</td>
<td>Decreases MMP-2, MMP-9, Smad-3, Snail-1, Twist-1 expression</td>
<td>Human renal cell carcinoma</td>
<td>112</td>
</tr>
<tr>
<td><strong>PAC</strong>*</td>
<td>Curcumin analogue</td>
<td>Inhibitor of EMT; antimigratory; antiproliferative</td>
<td>Inhibits JAK2/STAT3, AKT/mTOR and MEK/ERK signalling; inhibits STAT3/cyclin D1 pathway</td>
<td>Human colorectal cancer cells; tumour-bearing mice</td>
<td>113</td>
</tr>
<tr>
<td><strong>Palbociclib</strong></td>
<td>Selective CDK4/6 inhibitor</td>
<td>Inhibitor of EMT; antimigratory</td>
<td>Inhibits c-Jun/COX-2 signalling</td>
<td>Human breast cancer cells; tumour-bearing mice</td>
<td>114</td>
</tr>
<tr>
<td><strong>Pargyline</strong></td>
<td>Lysine-specific demethylase 1 inhibitor</td>
<td>Inhibitor of EMT; antimigratory</td>
<td>Reduces prostate-specific antigen</td>
<td>Human prostate cancer cells; tumour-bearing mice</td>
<td>115</td>
</tr>
<tr>
<td><strong>Physcion</strong></td>
<td>Anthraquinone derivative</td>
<td>Inhibitor of EMT; antimigratory</td>
<td>Activates ROS/AMPK/GSK3β signalling; inhibits SOX2</td>
<td>Human colorectal cancer cells</td>
<td>116</td>
</tr>
<tr>
<td><strong>Pterostilbene</strong>†</td>
<td>Antioxidant</td>
<td>Inhibitor of EMT; antimigratory</td>
<td>Reduces Src/Fak signalling; upregulates miR-205 expression</td>
<td>Human breast cancer cells; tumour-bearing mice</td>
<td>117</td>
</tr>
<tr>
<td><strong>Salidroside</strong>*</td>
<td>Glucopyranoside derivative</td>
<td>Inhibitor of EMT; antimigratory; antiproliferative</td>
<td>Downregulates miR-891b expression; inhibits PI3K/AKT/mTOR and NF-κB signalling</td>
<td>Wilms’ tumour cells; mouse prostate cancer</td>
<td>118</td>
</tr>
<tr>
<td><strong>Selenium</strong>†</td>
<td>Metal</td>
<td>Inhibitor of EMT</td>
<td>Downregulates genes involved in cellular migration, inflammation and mesenchymal markers; upregulates genes involved in epithelial markers</td>
<td>Human prostate</td>
<td>119</td>
</tr>
</tbody>
</table>

**Table 1** continued
beneficial effects on bleomycin-induced pulmonary fibrosis in rats by modulating the ACE2/ANG-(1–7) axis and by inhibiting lung inflammation.76 Selenium restores the antioxidant capacity of the lungs and reduces inflammatory responses, thus improving lung mechanics.77 All the previous examples concerned infection-induced lung injury, while lung injury may also be caused by the mechanical stress caused by forced ventilation. To specifically address this type of damage, Tsaknis et al.28 set up an elegant experimental model, exercising high-pressure ventilation on ex vivo lung preparations thereby increasing their microvascular permeability, oedema and microhaemorrhages; in this model, pretreatment with metformin decreased the severity of the damage preserving alveolar capillary permeability. Additional FDA-approved drugs that have already been used in clinical trials, but have not been discussed here, are metapristone, N-acetylcycteine, chrysin, berberine, salidroside, dehydroepiandrosterone and cisplatin. It is worth noting that many of the compounds above have been tested against cancer and reported to inhibit cell growth and EMT, but will not be discussed in this article. High-throughput screening in cell culture systems using a reporter system driven by the vimentin promoter will identify additional novel drugs for their capacity to specifically decrease vimentin transcription. In addition, alternative therapeutic interventions based on the downregulation of vimentin can be designed, such as the administration of antisense oligonucleotides, anti-vimentin RNA, humanised neutralising antibodies or dominant-negative peptides capable

A selection of compounds downregulating vimentin expression and their major cellular and molecular effects. Some of these molecules already have FDA approval, thus can be immediately used to set up clinical trials against COVID-19.

<table>
<thead>
<tr>
<th>Name of molecule</th>
<th>Type</th>
<th>Effects</th>
<th>Modes of action</th>
<th>Models</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Simvastatin††</td>
<td>Statin</td>
<td>Inhibitor of EMT; antifibrotic</td>
<td>Decreases Toll-like receptor 4 and NF-κB signalling</td>
<td>Human biliary epithelial cells</td>
<td>120</td>
</tr>
<tr>
<td>Sinomenine††</td>
<td>Alkaloid</td>
<td>Antimitragory; antiproliferative</td>
<td>Downregulates miR-23a expression; inhibits PI3K/AKT and JAK/STAT signalling</td>
<td>Human prostate cancer cells</td>
<td>121</td>
</tr>
<tr>
<td>Swainsoneine</td>
<td>Indolizidine alkaloid</td>
<td>Antimitragory; antiproliferative; proapoposis</td>
<td>Downregulates miR-92a, inhibits PI3K/AKT/mTOR signalling</td>
<td>Human glioblastoma cells</td>
<td>122</td>
</tr>
<tr>
<td>Tanshinone IIA††</td>
<td>Lipophilic compound</td>
<td>Inhibitor of EMT; antimitragory; antiproliferative</td>
<td>Inhibits STAT3-CCL2 signalling</td>
<td>Human bladder cancer cells</td>
<td>123</td>
</tr>
<tr>
<td>Tetramethylpyrazine nitro derivative</td>
<td>Tetramethylpyrazine derivative</td>
<td>Reduces brain infarction; preserves neurological function</td>
<td>Decrease expression of neuroinflammatory markers</td>
<td>Cynomolgus macaques brain ischaemic stroke model</td>
<td>124</td>
</tr>
<tr>
<td>Tetrandrine</td>
<td>Alkaloid</td>
<td>Inhibitor of EMT; antimitragory; antiproliferative</td>
<td>Reduces glioma-associated oncogene family zinc finger 1 (Gli-1) expression</td>
<td>Human bladder cancer cells</td>
<td>125</td>
</tr>
<tr>
<td>Toosendanin</td>
<td>Alkaloid</td>
<td>Inhibitor of EMT; antimitragory; antiproliferative</td>
<td>Inhibits AKT/mTOR signalling</td>
<td>Human pancreatic cancer cells; tumour-bearing mice</td>
<td>126</td>
</tr>
<tr>
<td>Triptolide††</td>
<td>Diterpenoid derivative</td>
<td>Antifibrotic</td>
<td>Inhibits TLR4-induced NF-κB/IL-1β immune pathway; inhibits NF-κB/TNF-α/VCAM-1 inflammatory pathway; downregulates TGF-β1/Smad3/vimentin fibrosis pathway</td>
<td>Diabetic rats</td>
<td>127</td>
</tr>
<tr>
<td>Valproic acid††</td>
<td>Acid</td>
<td>Inhibitor of EMT; antimitragory</td>
<td>Downregulates Smaad4 expression; upregulates transcriptional intermediary factor-1 (TIF1γ) expression</td>
<td>Human prostate carcinoma cells; tumour-bearing mice</td>
<td>128-129</td>
</tr>
<tr>
<td>Wogonin*</td>
<td>Flavonoid derivative</td>
<td>Inhibitor of EMT; antimitragory</td>
<td>Inhibits IL-6/STAT3 signalling pathway</td>
<td>Human alveolar adenocarcinoma cells</td>
<td>130</td>
</tr>
</tbody>
</table>

*Indicates the compounds have been referenced by the FDA; the latter group of drugs is briefly discussed in the text.
of blocking the interaction between vimentin and viral proteins.

CONCLUSIONS

Vimentin functions as a co-receptor for SARS-CoV—and likely for SARS-CoV-2—thus contributing to viral infection. Evidence of the significant role played by vimentin in virus-induced infection comes from the following observations: vimentin expression increases during viral infection in several clinical settings, while the absence of vimentin in knockout mice makes them more resistant to inflammation and acute lung injury than wild-type mice. Therefore, vimentin can be a target for the treatment of COVID-19-related pneumonia. It is a matter of fact that drugs that have proven efficient against viral infection downregulate vimentin expression. While the correlative nature of this evidence does not yet prove a causative role for vimentin downregulation in diminishing viral infection, it indicates that establishing whether drugs which decrease vimentin expression can be used for the treatment of patients with COVID-19 is the point at issue. Since vimentin has a twofold role in the disease, not only being involved in the viral infection but also in the associated life-threatening lung inflammation, the use of vimentin-targeted drugs may offer a synergistic advantage as compared with other treatments not targeting vimentin. In particular: (1) the decreased amount of vimentin on the cell surface would decrease its interaction with SARS-CoV-2 spike protein, thus interfering with viral entry; (2) the decreased amount of vimentin within the cell would affect virus replication; (3) the lesser amount of vimentin within inflammatory cells would exert negative effects on the inflammasome. This in turn would avoid the cytokine storm and curb the arrival of infiltrating inflammatory cells; (4) lower vimentin expression in epithelial cells and fibroblasts would decrease EMT and fibrosis. Taken together, all these considerations strongly suggest that vimentin represents a valuable therapeutic target at the early stages of the SARS-CoV infection, may help avoid the progression to serious complications and indicate that several FDA-approved drugs (such as melatonin, niclosamide, selenium, hydrogen sulfide, inositol) should be tested in clinical trials against SARS-CoV-2.

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ORCID iD  Ornith Agbolutu http://orcid.org/0000-0003-1923-8871

REFERENCES


28 Ziegler CG, Fallon SJ, Nyquist SK, et al. SARS-CoV-2 receptor ACE2 is an interferon- 

29 Grid the T


55 Li K, He Z, Wang X, et al. Apigenin C-glycosides of Microcos paniculata protects lipopolysaccharide induced apoptosis and...


